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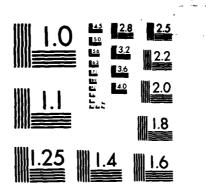
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Conditions for mercury deposition at cylindrical carbon fiber microelectrodes were examined and presented together with a new construction of electrode body and the manner in which the fiber is held. Data obtained in KSCN and acetate buffer solutions suggest that in order to obtain linear dependence of square wave stripping peak vs. mercury concentration the amount of the deposit should not exceed few monolayers. This could be caused by surface inhomogeneity as confirmed by electron microscopy and by the limited amount of mercury that can be (Continued)

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J. Golas and Janet Osteryoung*

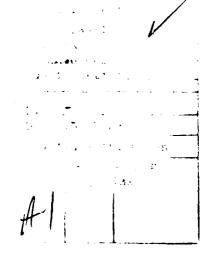
Department of Chemistry

SUNY at Buffalo

Buffalo, NY 14214

^aPermanent Address: Academy of Mining and Metallurgy, Institute of Material Science, 30*059 Krakow, Poland





Summary

Microelectrodes were examined and presented together with a new construction of electrode body and the manner in which the fiber is held. Data obtained in KSCN and acetate buffer solutions suggest that in order to obtain linear dependence of square wave stripping peak vs. mercury concentration the amount of the deposit should not exceed few monolayers. This could be caused by surface inhomogeneity as confirmed by electron microscopy and by the limited amount of mercury that can be quantitatively deposited at the surface. Several remarks on lead deposition "in situ" are also given.

Introduction

Several papers have been published recently on different types of carbon fiber microelectrodes [1-4]. Edmonds has reviewed this and related work in the context of the properties of the carbon fibers [5]. This material seems to indicate that carbon fibers can be used either directly as carbon electrodes or as a base material for mercury film electrodes. Although the results obtained with carbon fibers are still sometimes difficult to compare with each other, probably due to variability of the starting material and different pretreatment procedures, the increasing volume of experimental data followed by some theoretical considerations [6,7] is bringing carbon fibers closer to the point of general practical application. Three types of carbon fiber electrodes have been used: 1) the exposed single fiber with cylindrical geometry; 2) single fibers or arrays of fibers embedded in an insulator and having disk geometry; and 3) brushes, or arrays of exposed fibers. The first two seem to offer the best possibilities as voltammetric electrodes. Behavior of these electrodes depends on dimensions and shape and also on the properties of the material. Diffusion and convection are difficult to control or reproduce with the brush configuration [2].

Procedures for fabricating carbon fiber microelectrodes are improving [1,8,9], which should encourage and simplify further investigations. We present below an improvement over our previous procedure for making exposed carbon fiber microelectrodes [9]. The main aim of this paper is to indicate limitations of exposed carbon fibers for mercury deposition and for in situ deposition of lead. Relations between the properties of the apparent film of lead amalgam and the resulting cyclic voltammograms of lead are also presented.

Although relatively long (ca. 8-10 mm) cylindrical carbon fiber electrodes, which are the subject of this paper, at first sight may seem to be less practical, because fragile, than corresponding disk electrodes, they in fact have several advantages. They are much less sensitive to imperfect seal between electrode and insulator than are disk electrodes, because the integrated current density in the vicinity of the seal is a minute fraction of the total. Non-planar diffusion at an exposed fiber depends on the radius of the fiber rather than its length, so electrode area and properties of cylindrical diffusion can be controlled independently, again in contrast to the disk. In particular, the length of the fiber can be adjusted to give the desired total current for a given type of experiment. Finally, the exposed fiber electrodes make it possible to investigate the properties of the surface of the cylinder as an electrode, which might be different from those of the crossection.

Experimental Section

Instrumentation and Reagents

An IBM electrolytic cell was used along with a three electrode system in which carbon fiber, platinum and saturated calomel electrodes were used as working, counter and reference electrodes, respectively. Two voltammetric techniques were used: square wave voltammetry (SWV) and staircase voltammetry (SCV). All voltammetric measurements were performed with a computer-controlled potentiostat based on a Digital Equipment Corp. PDP=8/e minicomputer [10]. Some additional coulometric measurements were carried out using an EG&G PARC Model 173 Potentiostat and with Model 179 Digital Coulometer.

Reagents were of analytical grade and distilled water passed through a Millipore "Milli Q" purification system was used for preparation of the solutions. The following solutions were used as supporting electrolytes: 0.1 M KSCN (pH = 2.5; acidified with HNO_3), acetate buffer (1.25 M KOAc, 1.7 M HOAc, pH = 4.6), 0.1 M KNO_3 . The proper amounts of 0.05 M $Hg(NO_3)_2$ and 0.01 M $Pb(NO_3)_2$ were added to a chosen supporting electrolyte using Eppendorf pipets.

Solutions were purged with argon for about 7 min before each experiment; after purging the argon flow was directed over the solution.

All experiments were run in quiet solutions without stirring. According to our previous experience stirring during deposition causes the fiber to move which worsens reproducibility of stripping analysis.

Electrodes - Construction and Preliminary Tests

The working electrodes were made from 8 μ m-diameter carbon fibers (AESAR, Johnson Matthey Inc.). The procedure for making the electrode is based on an idea which involves sealing the fiber into a matrix of tygon and

heat-shrinkable tubing [7]. Gentle heating of the tip of the electrode with a soldering iron melts the inner tygon tube and shrinks the outer tube about the fiber and molten tygon. The main change from the procedure of Ref [7] is the Teflon body and glass support, as shown in Fig. 1. This body enables very quick replacement of the fiber in the case of damage and improves protection against breaking by providing the additional support which holds the tip of the fiber.

It should be noted that fibers even from the same manufacturer differ according to the smoothness of the surface. There are also different shapes of cross sections (e.g. AESAR fibers are circular whereas Celion GY-70 is bilobal). An electron—micrograph of AESAR carbon fibers is presented in Fig. 2. The surface is randomly covered with small dimples which did not disappear after pretreatment. The standard pretreatment procedure included cleaning in a solution of Alconox detergent in an ultrasonic bath, rinsing with water, cleaning again in HNO₃:H₂O (1:4) solution in the ultrasonic bath, rinsing with water, and finally drying. The last step was a moderate electrooxidation at +0.3 V for 1 min.

In principle, after cleaning one could select the proper part of the best-looking fiber by microscopic examination, but this procedure would be too time-consuming to yield practical electrodes. Furthermore, there is considerable uncertainty regarding what properties might be desirable to optimize performance in voltammetric experiments. A series of experiments were carried out on different fibers to examine the quantitative variability from fiber to fiber. Four carbon fibers were used for deposition of mercury and the resulting square wave anodic stripping voltammograms were compared for four deposition times (10, 20, 40, 100s), two deposition potentials (-0.6, -1.0 V) and several concentrations of

Hg(II) in the range 1#80 µM in 0.1 M KSCN. Although the results for each particular fiber are reproducible for deposition for a given time and concentration of Hg(II) (usually three repeated scans gave identical curves) different results were obtained for different fibers. For illustration representative data are presented in Table I. These electrochemical results suggest that the heterogeneous appearance of the surfaces shown in Fig. 2 in fact reveal differences in surface properties which affect electrochemical behavior.

Based on this preliminary series of experiments we chose one of those electrodes for further investigations. This was the carbon fiber microelectrode of length 9.06 mm (by microscopic measurement) and area of 0.228 mm². This electrode was then successfully used for several months without any damage. All of the following data were obtained using this one electrode.

Results and Discussion

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Mercury has been deposited on carbon fiber surfaces but it has not been established how much mercury can be deposited quantitatively. Following previous results [9] we investigated more carefully the conditions for mercury deposition in three different electrolytes: potassium thiocyanate, acetate buffer and potassium nitrate. The results in the first two supporting electrolytes are discussed together because they are qualitatively the same, whereas the nature of the mercury deposition in KNO_3 is a bit more complex. A series of depositions were run at $E_d = 41.0 \text{ V for}$ to $C_d = 10$, 20, 40 or 100s in the range of Hg(II) concentration from $C_d = 10$. The observed dependence of anodic stripping peak current density on mercury concentration in 0.1 M KSCN and in acetate buffer is shown in Figs. 3 and 4, respectively.

At sufficiently low concentration of Hg(II) or short deposition time, stripping peak current depends linearly on concentration of Hg(II), but at higher concentrations or longer times the dependence is less strong. This is more obvious for 0.1 M KSCN solution, where in general we obtained approximately three times larger stripping peaks than for acetate buffer under the same conditions. As a result in 0.1 KSCN the curves of Figure 3B become non-linear at values of concentration smaller than those of Figure 4B.

In order to estimate the amount of deposited mercury at which this effect starts, we measured charge consumed during each deposition. There is not a sharp boundary between the linear and non-linear ranges. We defined the boundary region as that in the range of conditions between 40 µM Hg(II) with 10s deposition and 20 µM Hg(II) with 100s deposition. The mean values for five replicate measurements were the following: $Q = 4.37 \mu C$ (S_x = 0.34 μ C) for [Hg(II)] = 20 μ M; Q = 0.99 μ C (S = 0.02 μ C) for [Hg(II)] = 40 μ M. It is well-known that deposits of mercury on carbon substrates are not coherent films but rather exist as collections of puddles or droplets [9]. We have observed such deposits on carbon fiber electrodes. However, bearing this in mind. it is useful to calculate the effective film thicknesses corresponding to these values of charge. Using L = 9.06 mm and r = 4 μm , the critical range of thickness of the deposit corresponding to these values of charge is 3414 A. Using a value of 1.44 A as the atomic radius of Hg, these numbers suggest that the stripping peak current depends linearly on coverage only up to a few monolayers.

At higher coverages reproducibility of the anodic stripping peak worsens, the peak becomes wider and more flat, and even the peak area (which should be proportional to total charge) does not depend linearily on

deposition time. The latter point is shown in Fig. 5. Examples were taken of two different concentrations of Hg(II) and two different deposition times. As is seen in Fig. 5, for values of $i_p w_{1/2}/A$ (peak current density times width at half height) in the range 5-10 μ A·V/mm² the dependence on deposition time decreases. This range of values again corresponds to an average thickness of several angstroms, agreeing with the results of Figs. 3 and 4. The same trends characterized by the same critical numbers occured in acetate buffer. Therefore this effect is due to the surface of the carbon fiber itself, and is not due only to the special characteristics of thiocyanate ion.

Similar experiments were performed in 0.1 M KNO2. Some typical anodic stripping voltammograms are shown in Fig 6. Peak current is linear with concentration of Hg(II) up to concentrations for which the second peak appears. But the currents are 10-20 times smaller than those obtained in thiocyanate or acetate solution. The results are qualitatively the same with added nitric acid (pH≥2.5). Clearly the deposition of mercury is hindered and the stripping process is complicated in this medium. The equilibrium constant for the reaction of ${\rm Hg}^{2+}$ with ${\rm Hg}$ to form ${\rm Hg}_2^{2+}$ is 88 [14]. Equilibrium for this reaction is established rapidly, so in the vicinity of the electrode when Hg is present the major form in solution should be Hg_2^{2+} . The anions SCN, $\mathrm{CH}_3\mathrm{COO}$, NO_3 all form insoluble salts of the composition Hg_2X_2 . In the case of SCN, the Hg(II) oxidation state is stabilized by formation of $Hg(SCN)_{ij}^{2-}$. The equilibrium constant for the reaction $Hg_2(SCN)_2(s) + 2SCN = Hg(SCN)_4^2 + Hg is 1 L/mol [14], and$ therefore in 0.1 M KSCN one would expect no precipitation of ${\rm Hg}_2({\rm SCN})_2({\rm s})$ in the presence of Hg for concentrations of Hg(II) less than 0.01 M. For acetate, again HgX_{ij}^{2-} is the predominant form, and the equilibrium constant

for the disproportionation reaction is sufficiently large that no precipitate should form [14-16]. Nitrate is quite different. The solubility product of $\mathrm{Hg}_2(\mathrm{NO}_3)_2 \cdot 2\mathrm{H}_2\mathrm{O}(s)$ estimated from the enthalpy values of Ref [14] is given by $\log K = -57 \; (\mathrm{mol/L})^3$. (It is "well-known", however that $\mathrm{Hg}_2(\mathrm{NO}_3)_2$ is quite soluble in slightly acid aqueous solution). Stabilization of $\mathrm{Hg}(\mathrm{II})$ by complex formation is negligible [17]. Therefore the undesirable behavior of mercury deposits in nitrate media appears due to the presence of $\mathrm{Hg}(\mathrm{I})$ and may reflect local precipitation of a form of $\mathrm{Hg}_2(\mathrm{NO}_3)_2$ in the vicinity of the electrode.

Anodic stripping of codeposited lead. The following series of experiments was carried out to test the range of conditions suitable for in situ codeposition of lead. The solution was 0.1 M KSCN (pH = 2.5) with [Hg(II)] = $40 \mu M$. Deposition was carried out without stirring for 60s at -0.6V. The nominal thickness of the mercury deposit under these conditions is 18 A. After deposition anodic square wave voltammograms were obtained with $E_{SW} = 25$ mV, $\Delta E_{g} = 6$ mV, and f = 100 Hz. The electrode was conditioned before each deposition for 1 min at +0.3 V. The procedure was repeated for varied concentrations of lead. Anodic stripping peak current increases linearly with concentration of lead up to 4 µM. This is shown in Fig. 7. However, at $[Pb(II)] = 1 \mu M$, the anodic stripping peak for Hg is diminished and shifted to more negative potential, and this effect is systematically more pronounced, the higher the concentration of Pb(II). An example is shown in Fig. 8. The solubility of Pb in Hg is 0.013 mol Pb/mol Hg. The smolar ratio of Pb to Hg in the amalgam under the conditions of this experiment, assuming deposition is diffusion-controlled, is just the concentration ratio, [Pb(II)]/[Hg(II)]. In the present case [Hg(II)] = 40 μ M, so saturation should occur for [Pb(II)] \geq 0.5 μ M. We observe the first

established that saturation must be avoided to obtain acceptable results for anodic stripping voltammetry [11-13]. In the present case the calibration curve remains linear up to a concentration of <u>ca</u> 8x the saturation value. For concentrations of Pb(II) greater than about 8 μ M (-15x the saturation value) we obtained broader anodic stripping peaks between -0.4 and -0.5 V and no peak for stripping of Hg. This effect has been reported previously for Cd [9].

Finally we would like to present few observations on the practical use of mercury coated carbon fiber microelectrodes. Fig. 9 presents two cyclic staircase voltammograms, both obtained in thiocyanate solution with Hg(II) concentration 40 µM and Pb(II) concentration 1.2 mM. Curve 1 (Fig 9) was obtained on a clean fiber, whereas curve 2 was obtained using a mercury-coated fiber (deposit thickness of ~95Å).

The voltammogram is improved dramatically in the presence of the mercury deposit. We have not found reports on experiments with similar conditions. Presently the theoretical basis for treating this problem, which involves aspects of cylindrical diffusion, nucleation and growth, and thin films, does not exist. However the practical implications for developing analytical methods suggest that this phenomenon should be examined in much greater detail.

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Conclusions

Although an exposed carbon fiber by itself is not convenient to operate with, the proposed construction of electrode body and the manner in which the fiber is held seems to ensure easy replacement of the fiber and makes the electrode relatively durable. Electron microscopic pictures of carbon fibers show a significant discrimination of the surface which could cause the differences in results while using different fibers. The applied pretreatment procedure did not affect significantly the surface properties.

According to our results, mercury can be deposited on carbon fibers, but stripping peak height depends linearly on mercury concentration only when the amount of deposit is less than a few monolayers. There are also some limitations for lead deposition "in situ". The best results are obtained for approximately 20A-thickness of the Hg deposit. The anodic stripping peak height increases linearly with lead concentration in the solution until the extent of supersaturation for the lead-mercury amalgam exceeds about seven times the value of saturation. The results obtained by using cyclic staircase voltammetry for lead on mercury-coated carbon fibers indicate that their quality is very sensitive to thickness of the mercury deposit. The best-shaped curves were obtained for deposits in the nominal thickness range of 50÷150A.

This result depends on the properties of the material, its dimensions, and possibly the vertical position of the electrode.

References

- 1. M. R. Cushman, B. G. Bennett and C. W. Anderson, Anal. Chim. Acta 130 (1981) 323.
- 2. G. Schultze and W. Frenzel, Anal. Chim. Acta 159 (1984) 95.
- 3. M. A. Dayton, J. C. Brown, K. J. Stutts and R. M. Wightman, Anal. Chem. 52 (1980) 946.
- 4. V. J. Jennings and J. E. Morgan, Analyst. 110 (1985) 121.
- 5. T. E. Edmonds, Anal. Chim. Acta, 175 (1985) 1-22.
- 6. K. Aoki, K. Honda, K. Tokuda and H. Matsuda J. Electroanal. Chem. 182 (1985) 267.
- 7. K. Aoki, K. Honda, V. Tokuda and H. Matsuda J. Electroanal. Chem. 186 (1985) 79.
- 8. J. L. Ponchon, R. Cespuglio, F. Gonon, M. Jouvet and J. F. Pujol, Anal. Chem., 51 (1979) 1483.
- 9. J. Golas and J. Osteryoung, in press.
- 10. T. R. Brumleve, J. J. O'Dea, R. A. Osteryoung and J. Osteryoung, Anal. Chem. 53 (1981) 7024706.
- 11. M. T. Kozlovskii, A. J. Zebreva and V. P. Gladyshev, Amalgamy i ikh Primienene, Alma-Ata, 1971.
- 12. Z. Stojek, B. Stepnik, Z. Kublik, J. Electroanal. Chem. 74 (1976) 277.
- 13. T. R. Copeland and R. K. Skogerboe, Anal. Chem., 46 (1974) 1257A.
- 14. "Standard Potentials in Aqueous Solution, A. J. Bard, R. Parsons, and J. Jordon, Eds.; International Union of Pure and Applied Chemistry, 1985. Marcel Dekker, New York.
- 15. "Stability Constants of Metal-ion Complexes: Part B. Organic Ligands. IUPAC Chemical Data Series-No. 22. D. D. Perrin, Ed. Pergamon, Oxford, 1979.
- 16. "Atlas of Metal-Ligand Equilibria in Aqueous Solution," J. Kragsten, Ellis Horwood, Chicester, 1978.
- 17. "Critical Stability Constants Vol. 4: Inorganic Complexes," R. M. Smith and A. E. Martell, Plenum, New York, 1976.

Peak current density for square wave anodic stripping voltammograms

| •• | | i _p /A, μA/mm ² | | | |
|--------------|-------------------|---------------------------------------|-------------|-------------|--------------------|
| | A,mm ² | 0.265 | 0.279 | 0.164 | 0.228 ^b |
| Ed.V | | | | | |
| -0. 6 | | 33.32 | 21.33 | 46.16 | 44.96 |
| -1.0 | | 132.23 | 75.98 | 153.35 | 116.01 |

^aSolution: 80 μ M Hg(II) in 0.1 M KSCN; deposition time: 40 s; square wave parameters: Δ E = 4 mV, E = 25 mV, f = 100 Hz. The four different electrodes are identified by their areas.

Table I

^bThis electrode taken for further investigations.

Figure Captions

- Fig. 1. Schematic drawing of the cylindrical carbon fiber microelectrode.
- Fig. 2. Electron-microscopic picture of a bunch of the AESAR carbon fibers.
- Fig. 3. A. Anodic stripping square wave voltammograms of mecury in 0.1 M KSCN, pH = 2.5. Square wave parameters: $E_{gw} = 24$ mV, $\Delta E_{g} = 4$ mV, f = 100 Hz. $E_{d} = 41.0$ V, $t_{d} = 40$ sec. [Hg(II)]: 1) 8, 2) 20, 3) 40, 4) 80 µM.
 - B. Dependence of anodic stripping square wave peak current density on Hg(II) concentration for varied deposition time. $t_d(s) = (o) 10$, (A) 20, (A) 40, (e) 100. Other conditions those of Figure 3A.
- Fig. 4. A. Anodic stripping square wave voltammograms of mercury in 1.25/1.7 M acetate buffer pH = 4.6. [Hg(II)]: 1) 20, 2) 40, 3) 80 µM. Other conditions as in Figure 3A.
 - B. Dependence of anodic stripping square wave peak current density on mercury concentration for varied deposition time: (0) 10, (Δ) 20, (Δ) 40, (\oplus) 100 s.
- Fig. 5. Anodic stripping square wave voltammetry of mercury in 0.1 M KSCN. The dependence of peak area on deposition time. [Hg(II)] = 20 μ M, E_d = +0.6 V (Δ) or 41.0 V (Δ); [Hg(II)] = 80 μ m, E_d = =0.6 V (Δ) or A1.0 V (Δ). Peak area approximated as height times width at half height, $\mu_{1/2}$.

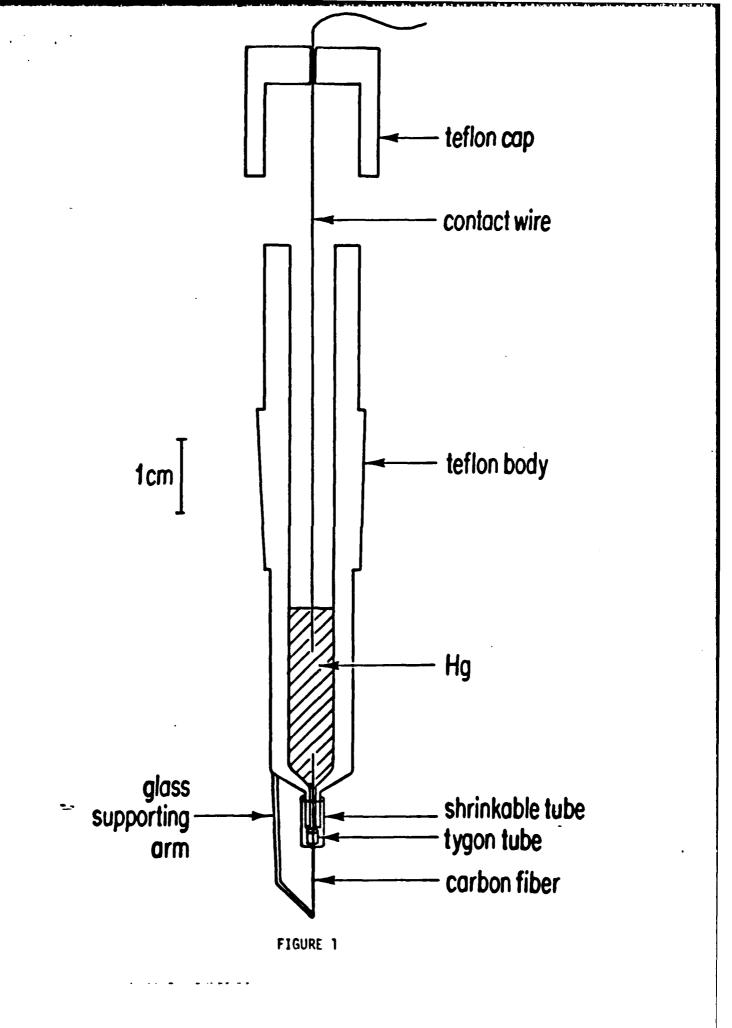
- Fig. 6. Anodic stripping square wave voltammograms of mercury in 0.1 M $\rm KNO_3$ [Hg(II)] (μM): 1) 8, 2) 20, 3) 40 (Fig. 6A) and 160 (Fig. 6B). Other conditions as Fig. 3A.
- Fig. 7. Anodic stripping square wave voltammetry of lead codeposited with mercury. Dependence of peak height on concentration of lead.

 (E_d = #0.6V; t_d = 60 s; SWV: E_{sw} = 24 mV, ΔE_s = 4 mV, f = 100 Hz;

 0.1 M KSCN, pH = 2.5. [Hg(II)] = 40 μM. Straight line fitted to linear portion; correlation coefficient = 0.9992.
- Fig. 8. Anodic stripping square wave voltammograms of mercury (curve 1) and mercury/lead amalgam (2). [Pb(II)] = 2.5 μ M. Other conditions as Fig. 7.
- Fig. 9. Cyclic staircase voltammograms (sweep rate 0.8 V/sec) Solution:

 0.1 M KSCN, pH = 2.5, [Hg(II)] = 0.04 mM, [Pb(II)] = 1.2 mM.

 Curve 1 without previous mercury deposition. Curve 2 mercury deposited at 40.3V. Apparent thickness of the deposit 2 95 Å.



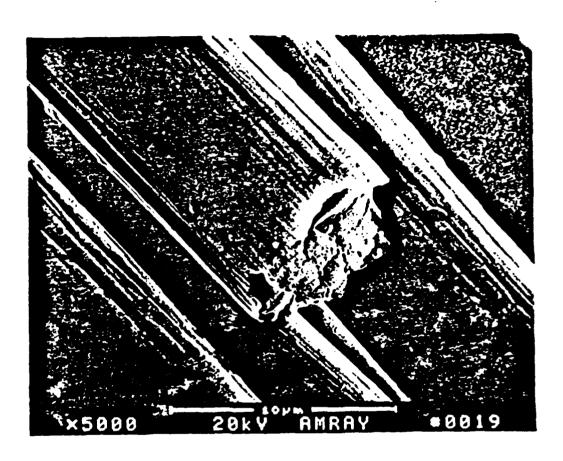
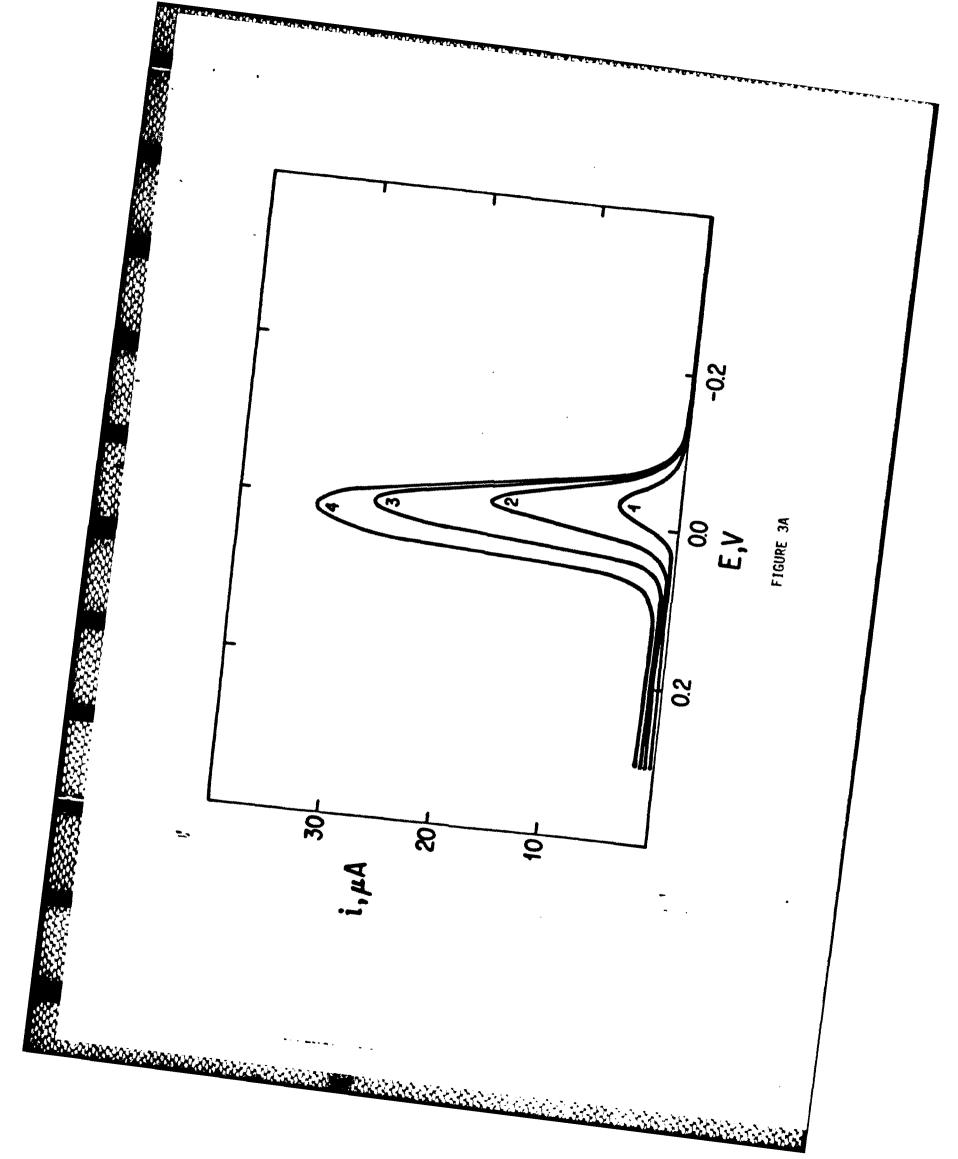
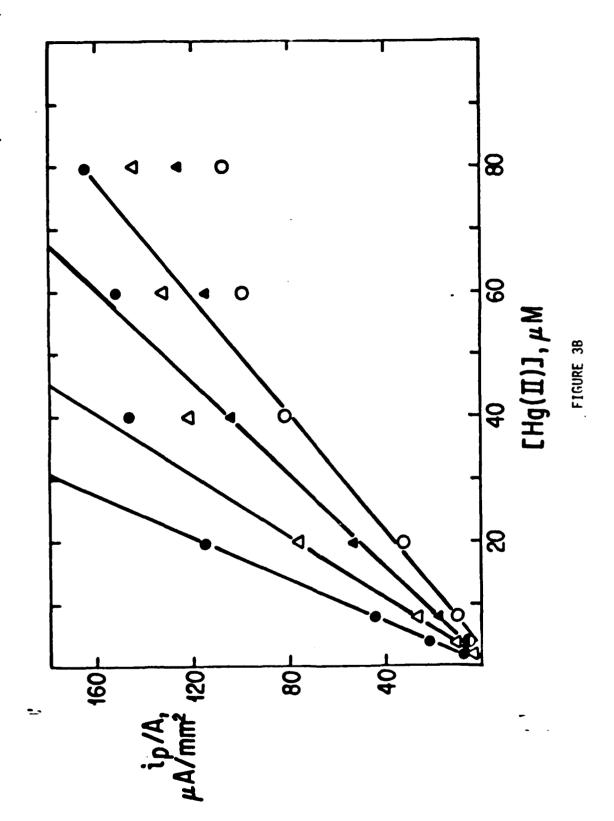
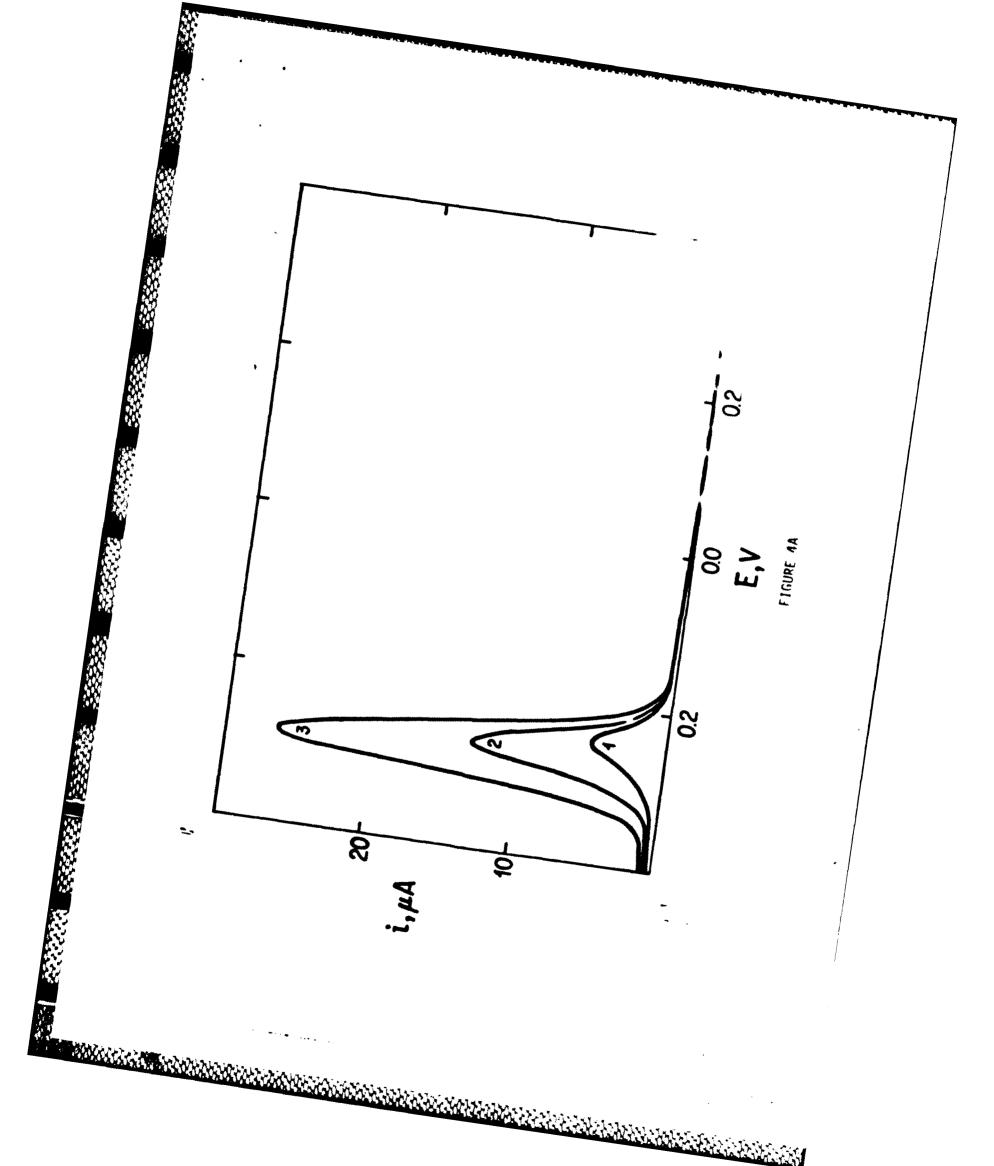
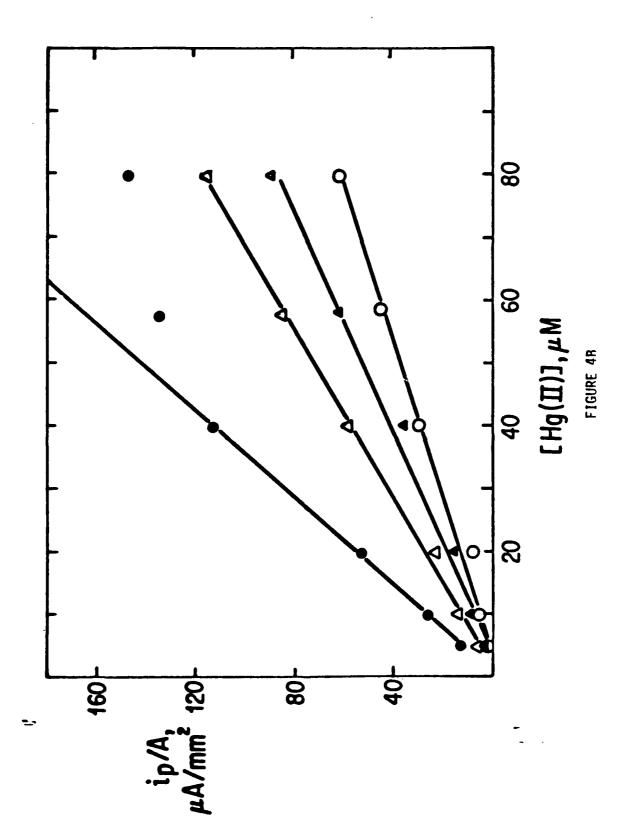


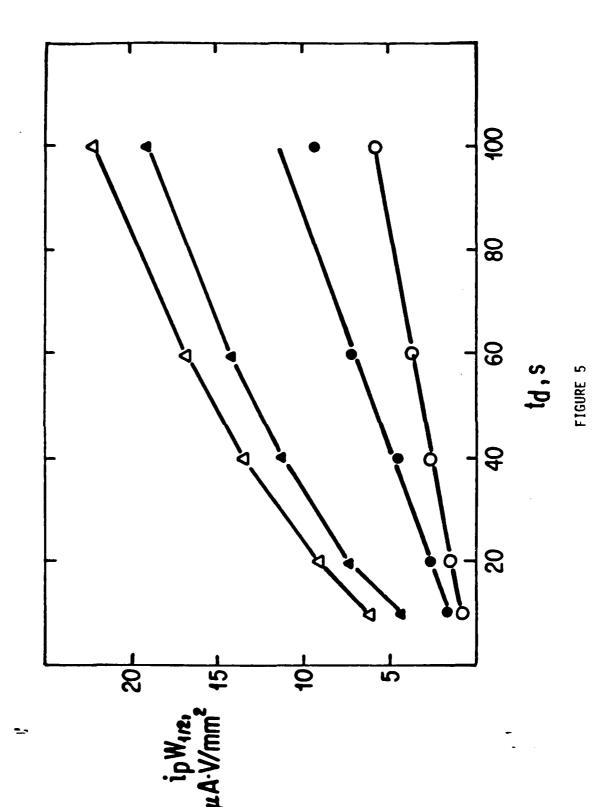
FIGURE 2

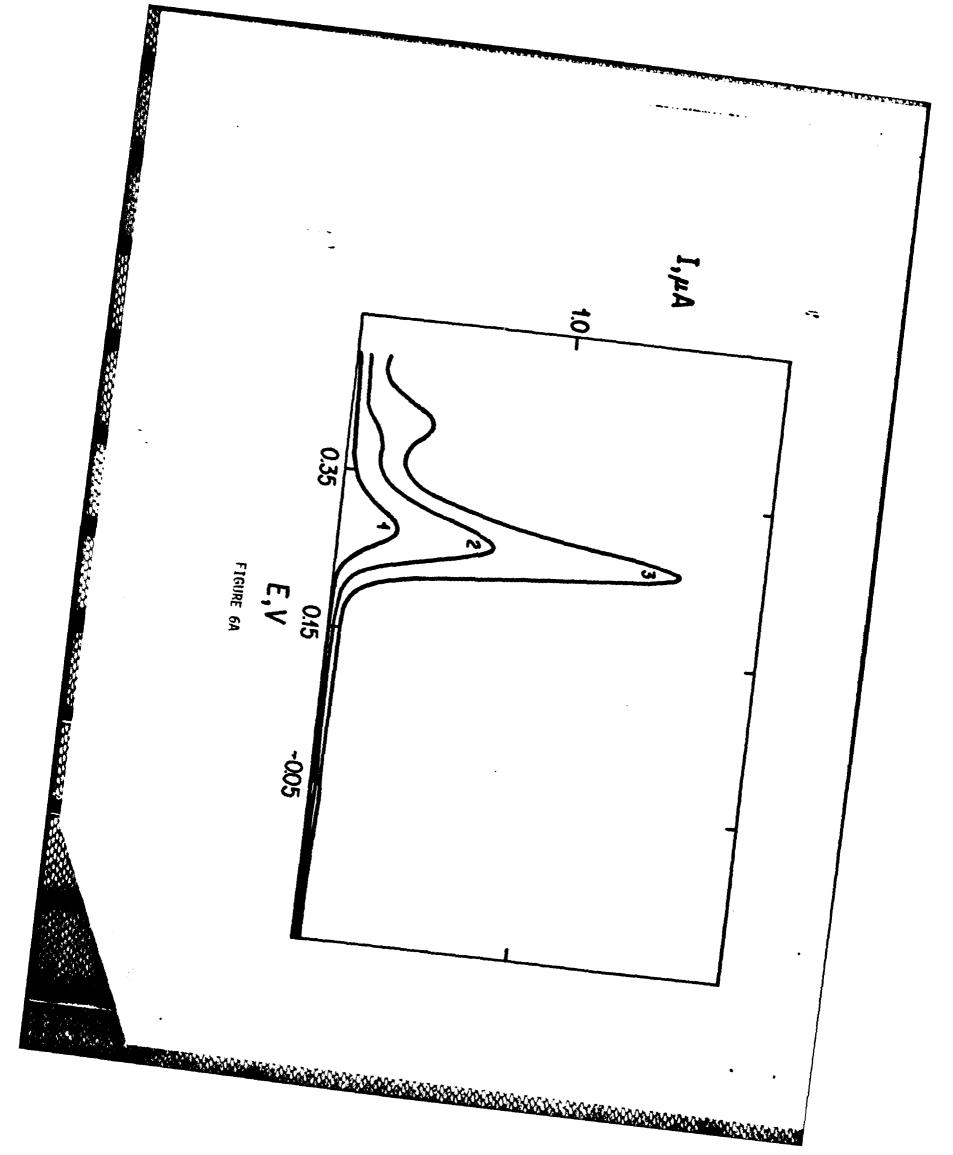


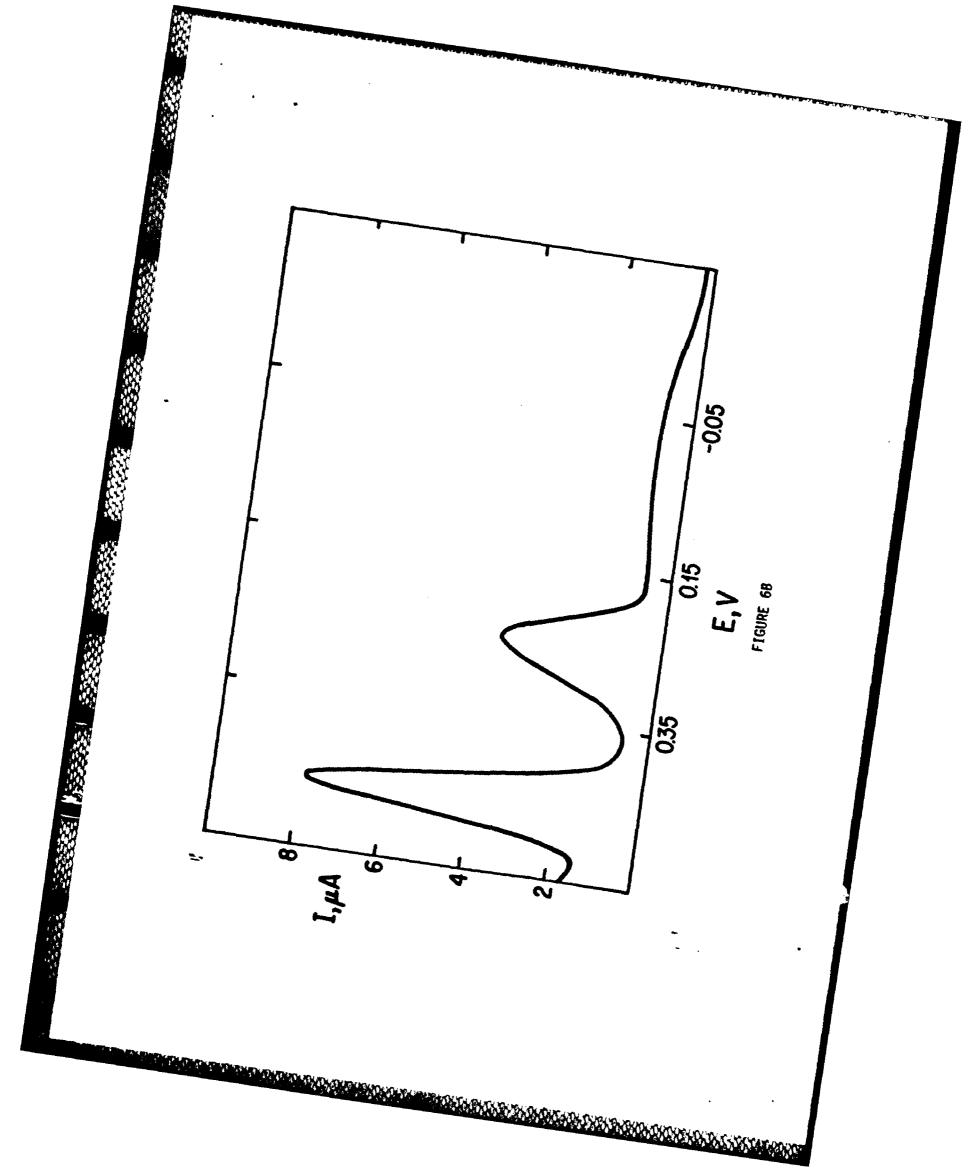


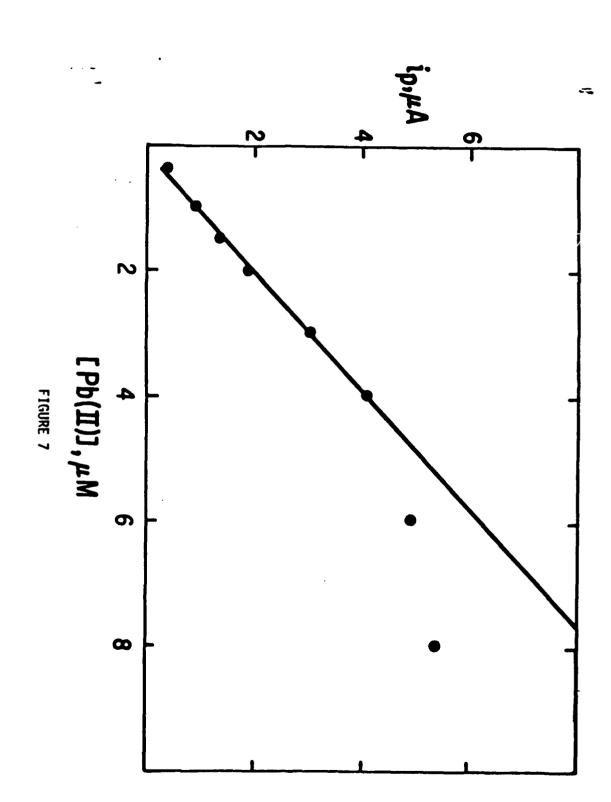


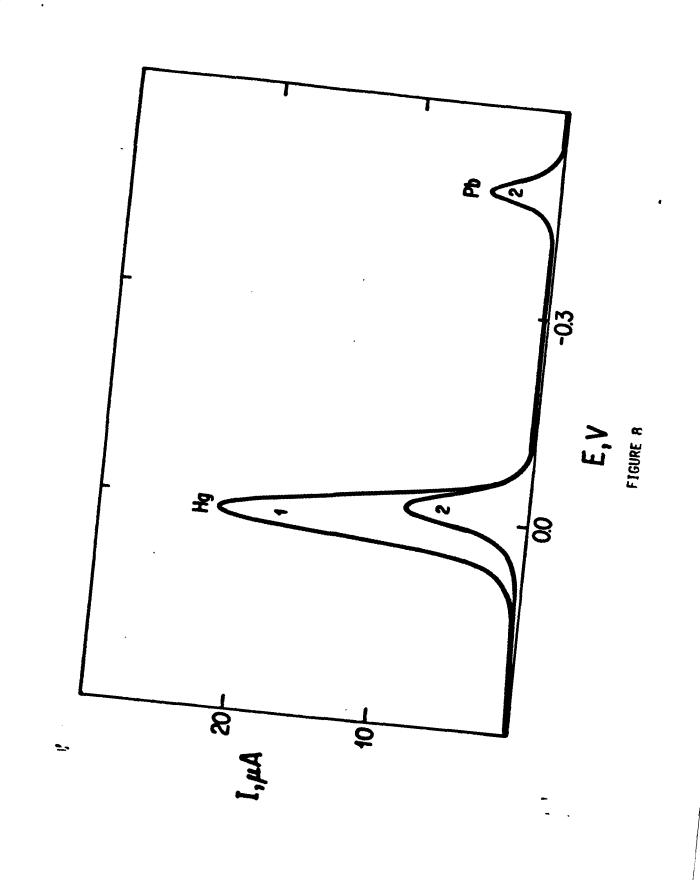


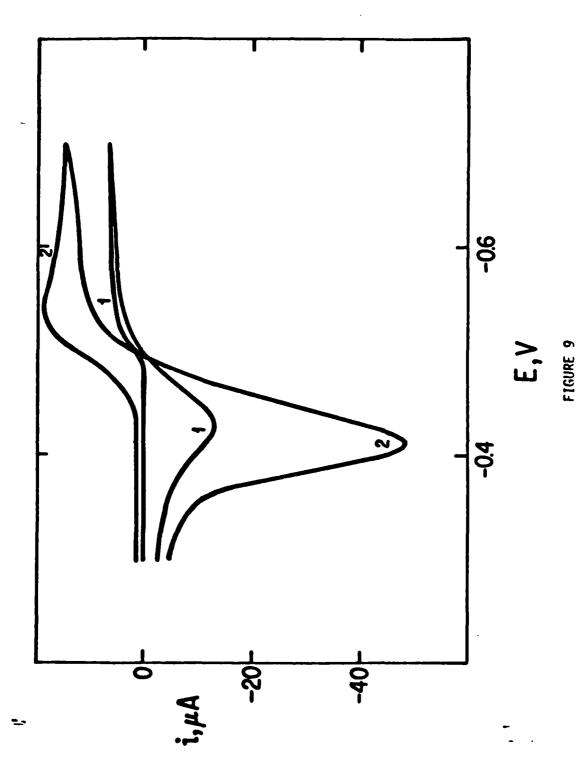












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Department of Chemistry
State University of New York
Buffalo, New York 14214

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Dr. S. P. Perone Lawrence Livermore Mational Laboratory L-370 P.O. Box 808 Livermore, California 94550 Dr. G. M. Hieftje Department of Chemistry Indiana University Bloomington, Indiana 47401

Dr. Christie G. Eake Department of Chemistry Michigan State University East Lansing, Michigan 48824

Walter G. Cox, Code 3632 Naval Underwater Systems Center Building 148 Newport, Rhode Island 02840

Professor Isiah M. Warner Department of Chemistry Emory University Atlanta, Georgia 30322

Dr. Kent Eisentraut Air Force Materials Laboratory Wright-Patterson AFB, Ohio 45433

Dr. Adolph B. Amster Chemistry Division Naval Weapons Center China Lake, California 93555

Dr. B. E. Douda Chemical Sciences Branch Code 50 C Naval Weapons Support Center Crane, Indiana 47322

Dr. John Eyler Department of Chemistry University of Florida Gainesville, Florida 32611

ABSTRACTS DISTRIBUTION LIST, 051B

Professor J. Janata
Department of Bioengineering
University of Utah
Salt Lake City, Utah 84112

Dr. J. DeCorpo NAVSEA Code 05R14 Washington, D.C. 20362

Dr. Charles Anderson Analytical Chemistry Division Athens Environmental Laboratory College Station Road Athens, Georgia 30613

Dr. Ron Flemming B 108 Reactor National Bureau of Standards Washington, D.C. 20234

Dr. Frank Herr Office of Naval Research Code 422CB 800 N. Quincy Street Arlington, Virginia 22217

Professor E. Keating
Department of Mechanical Engineering
U.S. Naval Academy
Annapolis, Maryland 21401

Dr. M. H. Miller 1133 Hampton Road Route 4 U.S. Naval Academy Annapolis, Maryland 21401

Dr. Clifford Spiegelman National Bureau of Standards Room A337 Bldg. 101 Washington, D.C. 20234 Dr. Denton Elliott AFOSR/NC Bolling AFB Washington, D.C. 20362

Dr. B. E. Spielvogel Inorganic and Analytical Branch P.O. Box 12211 Research Triangle Park, NC 27709

Ms. Ann De Witt Material Science Department 160 Fieldcrest Avenue Raritan Center Edison. New Jersey 08818

Dr. A. Harvey Code 6110 Naval Research Laboratory Washington, D.C. 20375

Mr. S. M. Hurley Naval Facilities Engineering Command Code 032P 200 Stovall Street Alexandria, Virginia 22331

Ms. W. Parkhurst Naval Surface Weapons Center Code R33 Silver Spring, Maryland 20910

Dr. M. Robertson Electrochemical Power Sources Division Code 305 Naval Weapons Support Center Crane, Indiana 47522

Dr. Andrew T. Zander PI204 Perkin-Elmer Corporation 901 Ethan Allen Highway/MS905 Ridgefield, Connecticut 06877

ABSTRACTS DISTRIBUTION LIST, 051B

Dr. Marvin Wilkerson Naval Weapons Support Center Code 30511 Crane, Indiana 47522

Dr. J. Wyatt Naval Research Laboratory Code 6110 Washington, D.C. 20375

Dr. J. MacDonald Code 6110 Naval Research Laboratory Washington, D.C. 20375 Dr. H. Wohltjen Naval Research Laboratory Code 6170 Washington, D.C. 20375

Dr. John Hoffsommer Naval Surface Weapons Center Building 30 Room 208 Silver Spring, Maryland 20910

Dr. Robert W. Shaw U.S. Army Research Office Box 12211 Research Triangle Park, NC 27709

ABSTRACTS DISTRIBUTION LIST, 359/627

Dr. Paul Delahay Department of Chemistry New York University New York, New York 10003

Dr. P. J. Hendra Department of Chemistry University of Southampton Southampton SO9 5NH United Kingdom

Dr. T. Katan
Lockheed Missiles and
Space Co., Inc.
P.O. Box 504
Sunnyvale, California 94088

Dr. D. N. Bennion
Department of Chemical Engineering
Brigham Young University
Provo, Utah 84602

Mr. Joseph McCartney Code 7121 Naval Ocean Systems Center San Diego, California 92152

Dr. J. J. Auborn
Bell Laboratories
Murray Hill, New Jersey 07974

Dr. Joseph Singer, Code 302-1 NASA-Lewis 21000 Brookpark Road Cleveland, Ohio 44135

Dr. P. P. Schmidt
Department of Chemistry
Oakland University
Rochester, Michigan 48063

SOCOOD RANKER CONTRACT SECOND

Dr. H. Richtol Chemistry Department Rensselaer Polytechnic Institute Troy. New York 12181

Dr. R. A. Marcus Department of Chemistry California Institute of Technology Pasadena, California 91125 Dr. E. Yeager Department of Chemistry Case Western Reserve University Cleveland, Ohio 44106

Dr. C. E. Mueller
The Electrochemistry Branch
Naval Surface Weapons Center
White Oak Laboratory
Silver Spring, Maryland 20910

Dr. Sam Perone
Chemistry & Materials
Science Department
Lawrence Livermore National Laboratory
Livermore, California 94550

Dr. Royce W. Murray
Department of Chemistry
University of North Carolina
Chapel Hill, North Carolina 27514

Dr. B. Brummer
EIC Incorporated
111 Downey Street
Norwood, Massachusetts 02062

Dr. Adam Heller Bell Laboratories Murray Hill, New Jersey 07974

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Dr. A. B. Ellis Chemistry Department University of Wisconsin Madison, Wisconsin 53706

Dr. Manfred Breiter
Institut fur Technische Elektrochemie
Technischen Universität Wien
9 Getreidemarkt, 1160Wien
AUSTRIA

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ABSTRACTS DISTRIBUTION LIST, 359/627

Dr. M. Wrighton
Chem'stry Department
Massachusetts Institute
of Technology
Cambridge, Massachusetts 02139

Dr. B. Stanley Pons
Department of Chemistry
University of Utah
Salt Lake City, Utah 84112

Donald E. Mains
Naval Weapons Support Center
Electrochemical Power Sources Division
Crane, Indiana 47522

S. Ruby
DOE (STOR)
M.S. 68025 Forrestal Bldg.
Washington, D.C. 20595

Dr. A. J. Bard
Department of Chemistry
University of Texas
Austin, Texas 78712

Dr. Janet Osteryoung
Department of Chemistry
State University of New York
Buffalo, New York 14214

Dr. Donald W. Ernst Naval Surface Weapons Center Code R-33 White Oak Laboratory Silver Spring, Maryland 20910

Mr. James R. Moden
Naval Underwater Systems Center
Code 3632
Newport, Rhode Island 02840

Or. Bernard Spielvogel
U.S. Army Research Office
P.O. Box 12211
Research Triangle Park, NC 27709

Dr. Aaron Fletcher Naval Weapons Center Code 3852 China Lake, California 93555 Dr. M. M. Nicholson Electronics Research Center Rockwell International 3370 Miraloma Avenue Anaheim, California

Dr. Michael J. Weaver
Department of Chemistry
Purdue University
West Lafayette, Indiana 47907

Dr. R. David Rauh EIC Laboratories, Inc. 111 Downey Street Norwood, Massachusetts 02062

Dr. Aaron Wold Department of Chemistry Brown University Providence, Rhode Island 02192

Dr. Martin Fleischmann Department of Chemistry University of Southampton Southampton SO9 5NH ENGLAND

Dr. R. A. Osteryoung Department of Chemistry State University of New York Buffalo, New York 14214

Dr. Denton Elliott
Air Force Office of Scientific
Research
Bolling AFB
Washington, D.C. 20332

Dr. R. Nowak Naval Research Laboratory Code 6170 Washington, D.C. 20375

Dr. D. F. Shriver
Department of Chemistry
Northwestern University
Evanston, Illinois 60201

Dr. Boris Cahan Department of Chemistry Case Western Reserve University Cleveland, Ohio 44106

ABSTRACTS DISTRIBUTION LIST, 359/627

Dr. David Aikens Chemistry Department Rensselaer Polytechnic Institute Troy, New York 12181

Dr. A. B. P. Lever Chemistry Department York University Downsview, Ontario M3J1P3

Dr. Stanislaw Szpak Naval Ocean Systems Center Code 6343, Bayside San Diego, California 95152

Dr. Gregory Farrington
Department of Materials Science
and Engineering
University of Pennsylvania
Philadelphia, Pennsylvania 19104

M. L. Robertson

Manager, Electrochemical
and Power Sources Division

Naval Weapons Support Center

Crane, Indiana 47522

Dr. T. Marks
Department of Chemistry
Northwestern University
Evanston, Illinois 60201

Dr. Micha Tomkiewicz Department of Physics Brooklyn College Brooklyn, New York 11210

Dr. Lesser Blum
Department of Physics
University of Puerto Rico
Rio Piedras, Puerto Rico 00931

Dr. Joseph Gordon, II IBM Corporation K33/281 5600 Cottle Road San Jose, California 95193

Dr. Hector D. Abruna
Department of Chemistry
Cornell University
Ithaca, New York 14853

Dr. D. H. Whitmore
Department of Materials Science
Northwestern University
Evanston, Illinois 60201

Dr. Alan Bewick Department of Chemistry The University of Southampton Southampton, SO9 5NH ENGLAND

Dr. E. Anderson NAVSEA-56Z33 NC #4 2541 Jefferson Davis Highway Arlington, Virginia 20362

Dr. Bruce Dunn
Department of Engineering &
Applied Science
University of California
Los Angeles, California 90024

Dr. Elton Cairns
Energy & Environment Division
Lawrence Berkeley Laboratory
University of California
Berkeley, California 94720

Dr. D. Cipris
Allied Corporation
P.O. Box 3000R
Morristown, New Jersey 07960

Dr. M. Philpott IBM Corporation 5600 Cottle Road San Jose, California 95193

Dr. Donald Sandstrom Boeing Aerospace Co. P.O. Box 3999 Seattle, Washington 98124

Dr. Carl Kannewurf
Department of Electrical Engineering
and Computer Science
Northwestern University
Evanston, Illinois 60201

Dr. Richard Pollard
Department of Chemical Engineering
University of Houston
4800 Calhoun Blvd.
Houston, Texas 77004

ABSTRACTS DISTRIBUTION LIST, 359/627

Dr. Robert Somoano
Jet Propulsion Laboratory
California Institute of Technology
Pasadena, California 91103

Dr. Johann A. Joebstl
USA Mobility Equipment R&D Command
DRDME-EC
Fort Belvoir, Virginia 22060

Dr. Judith H. Ambrus NASA Headquarters M.S. RTS-6 Washington, D.C. 20546

Dr. Albert R. Landgrebe U.S. Department of Energy M.S. 68025 Forrestal Building Washington, D.C. 20595

Dr. J. Brophy
Department of Physics
University of Utah
Salt Lake City, Utah 84112

Dr. Charles Martin
Department of Chemistry
Texas A&M University
College Station. Texas 77843

Dr. H. Tachikawa Department of Chemistry Jackson State University Jackson, Mississippi 39217

Dr. Theodore Beck Electrochemical Technology Corp. 3935 Leary Way N.W. Seattle, Washington 98107

Dr. Farrell Lytle
Boeing-Engineering and
Construction Engineers
P.O. Box 3707
Seattle, Washington 98124

Dr. Robert Gotscholl U.S. Department of Energy MS G-226 Washington, D.C. 20545 Dr. Edward Fletcher Department of Mechanical Engineering University of Minnesota Minneapolis, Minnesota 55455

Dr. John Fontanella Department of Physics U.S. Naval Academy Annapolis, Maryland 21402

Dr. Martha Greenblatt
Department of Chemistry
Rutgers University
New Brunswick, New Jersey 08903

Dr. John Wasson Syntheco, Inc. Rte 6 - Industrial Pike Road Gastonia. North Carolina 28052

Dr. Walter Roth
Department of Physics
State University of New York
Albany, New York 12222

Dr. Anthony Sammells Eltron Research Inc. 4260 Westbrook Drive, Suite 111 Aurora, Illinois 60505

Dr. W. M. Risen Department of Chemistry Brown University Providence, Rhode Island 02192

Dr. C. A. Angell Department of Chemistry Purdue University West Lafayette, Indiana 47907

Dr. Thomas Davis
Polymer Science and Standards
Division
National Bureau of Standards
Washington, D.C. 20234

Ms. Wendy Parkhurst Naval Surface Weapons Center R-33 Silver Spring, Maryland 20910